Optical tweezers for *in vivo* **DNA manipulation**

Vadim Bogatyr, Andreas Biebricher, Gijs Wuite

I. Vision

Creating an artificial cell, we strive to characterize and understand it as deeply as possible. Utilizing optical tweezers integrated with fluorescence spectroscopy and microfluidic techniques, I aim to build a system for in vivo manipulation of DNA. It will allow for a quantitative description of protein-DNA interaction inside the artificial cell during such fundamental processes of molecular biology as replication and transcription.



II. Optical traps in single-molecule DNA studies



Studying DNA anaphase ultra-fine bridges by three-color imaging of associated protein factors



monitored using STED nanoscopy TFAM diffusion on a dsDNA Confocal vs STED

Protein dynamics on a densly covered DNA



DNA bridging by XRCC4 and XLF investigated on a quadro-optical trap setup



IV. Confinement of the liposomes

If the liposomes are freely floating in the solution during the experiment, they will get trapped. This would obstruct successful bead manipulation. Possible solutions to this problem are:

Immobilization

Trapping

Deformation









III. Production of the GUVs



Inverse emulsion

Water-in-oil emulsion droplets are pushed through the lipid monolayer formed on the interface of lipid-in-oil and aqueous solutions to produce liposomes.



cDICE

An improved version of the inverse emulsion. Here, the water-in-oil droplets are produced with a capillary. The encapsulation efficiency and size monodispersity are higher.

V. Research roadmap



In vivo DNA measurements with optical tweezers

VI. Results and outlook



The inverse emulsion has a high yield and was already used for beads encapsulation (bottom) figure). Nevertheless, **cDICE** is the most prospective candidate for GUV production due to its fast



OLA



A microfluidics-based approach, where the double emulsion droplets of equally small size are flow focused. The dewetting process leads to the separation of the oil droplet from the liposome.

automated liposome production rates, high encapsulation efficiency and a lower degree of shear stress exerted on the inner solution. The latter is particularly important to prevent DNA damage.

Hoekstra, T. P. et al. Switching between Exonucleolysis and Replication by T7 DNA Polymerase Ensures High Fidelity. Biophys. J. 112, 575–583 (2017). Heller, I. et al. STED nanoscopy combined with optical tweezers reveals protein dynamics on densely covered DNA. (2013). Sarlós, K. et al. Reconstitution of anaphase DNA bridge recognition and disjunction. Nat. Struct. Mol. Biol. 25, 868–876 (2018). Brouwer, I. et al. Sliding sleeves of XRCC4-XLF bridge DNA and connect fragments of broken DNA. Nature 535, 566–569 (2016). Mulla, Y., Aufderhorst-Roberts, A. & Koenderink, G. H. Shaping up synthetic cells. Physical Biology vol. 15 (2018). Deshpande, S., Caspi, Y., Meijering, A. E. C. & Dekker, C. Octanol-assisted liposome assembly on chip. Nat. Commun. 7, (2016). Fanalista, F. et al. Shape and Size Control of Artificial Cells for Bottom-Up Biology. ACS Nano 13, 5439–5450 (2019). Robinson, T., Kuhn, P., Eyer, K. & Dittrich, P. S. Microfluidic trapping of giant unilamellar vesicles to study transport through a membrane pore. Biomicrofluidics 7, (2013). Ganzinger, K. A. et al. Microfluidic trapping of vesicles reveals membrane-tension dependent FtsZ cytoskeletal re-organisation. bioRxiv 791459 (2019).

