# S6 unfolds in a complex multi-pathway manner in the detergent SDS as opposed to simple two-state denaturation in GdmCl

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S6 - a model soluble protein for unfolding studies

In this work, the effects of chemical denaturant guanidinium chloride (GdmCl) and anionic detergent sodium dodecyl sulfate (SDS) on a model soluble protein S6<sup>1</sup> are compared and the unfolding pathways are disentangled on a single-molecule level.

## Protien denaturation by chemical denaturants and detergents

Chemical denaturants (GdmCl) and detergents (SDS) induce **drastically different denaturation modes** in proteins. The table below summarizes their main characteristics:

Single-molecule FRET reveals two-state unfolding of S6 in GdmCl



Major structural changes happen to S6 at sub-cmc SDS concentration

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![](_page_0_Figure_13.jpeg)

(A) Two distinct peaks correspond to the folded and unfolded states. The unfolding midpoint was found to be 3.2 M GdmCl

(B) FRET-2CDE analysis<sup>6</sup> assigned scores <20 to the majority of the observed S6 molecules. This indicated that they were **static** on the timescale of diffusion through the confocal volume (~ 1 ms). Previously reported unfolding rate of 0.032 s<sup>-1</sup> at the midpoint of 3.25 M GdmCl<sup>7</sup> is in great agreement with these results.

![](_page_0_Figure_16.jpeg)

![](_page_0_Figure_17.jpeg)

Expansion of S6 happens at 0.4 mM SDS (dotted line), cmc of SDS is 1.1 mM (dashed line).
(A) Hindrance of fluorophore rotation upon expansion indicated numerous SDS monomers associate with S6 and form a hemi-micelle-protein complex at sub-cmc SDS.
(B) Increase of acceptor lifetime was linked to the further spacial separation from Trp62, meaning that S6 was in a more expanded conformation in comparison to the native state.
(C) Blue shift of the Trp fluorescence spectra suggested a more hydraphobic microenvironment of the aa, i.e., the Trp is associated with alkyl tails of SDS, embeded within micelle. Thus, the final unfolded state at hundreds mM SDS is most likely necklace-and-beads<sup>8</sup>.
(D) In contrast to the random coil S6 in GdmCl, SDS promotes an α-helice-rich state<sup>9-10</sup>.
(E) Major loss of β-sheets occurs at sub-cmc SDS.

### Disentangling the heterogeneous populations of S6 below cmc with smFRET analysis toolkit

![](_page_0_Figure_20.jpeg)

(A) Heterogeneous extended states emerged around the cmc - unfolded ensemble.
(B) Their reconfiguration timescale, is much slower than the observation one.
(C) FRET histograms at 0.4 mM SDS evolve over minutes.

![](_page_0_Figure_22.jpeg)

To this end a custom-built multiparameter fluorescence detection (MFD) setup<sup>4</sup> was utilized to disentangle the conformational dynamics induced in S6 during the unfolding with the nanoscale temporal and spatial resolution<sup>5</sup>.

#### Experiments

#### Single-molecule FRET experiments:

S6 variant M1C/F97C containing two cysteine residues at positions 1 and 97 was engineered and stochastically labeled with FRET donor Atto 532 (Atto-Tec, Germany) and acceptor Abberior Star 635P (Abberior GmbH, Germany) fluorophores. Measurements were performed at 50 pM S6 concentration; (D) Unfolding at sub-cmc SDS occurs through a transient intermediate state.

(E) The kinetic rates of expansion are highly dependent on the SDS concentration and increase with addition of the detergent.

## S6 interconverts between different protein-micelle complexes with millisecond kinetics at high micellar density

![](_page_0_Figure_30.jpeg)

### Multiple pathways of S6 unfolding in SDS

![](_page_0_Figure_32.jpeg)

**Circular dichroism experiments:** 17 μM unlabeled M1C/F97C S6 variant;

Tryptophan fluorescence: 2  $\mu$ M of wt S6;

Condition: 150 NaCl, 50 mM Tris 7.4 pH, 2 mM TCEP;

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#### **References:**

M. B. Garber et al. Biochimie, vol. 74, no. 4, pp. 327–336, Apr. 1992.
 D. Otzen. Biochim. Biophys. Acta - Proteins Proteomics, vol. 1814, no. 5, pp. 562–591, 2011.
 C. Tanford et al. J. Biol. Chem., vol. 241, no. 8, pp. 1921–3, Apr. 1966.
 E. Sisamakis et al. Methods Enzymol., vol. 475, pp. 455–514, Jan. 2010.
 V. Kudryavtsev et al. ChemPhysChem, vol. 13, no. 4, pp. 1060–1078, Mar. 2012.
 T. E. Tomov et al. Biophys. J., vol. 102, no. 5, pp. 1163–1173, Mar. 2012.
 D. E. Otzen et al. Biochemistry, vol. 2, no. 38, pp. 6499–6511, 1999.
 M. Samsó et al. Eur. J. Biochem., vol. 232, no. 3, pp. 818–24, Sep. 1995.
 W. L. Mattice et al. Biochemistry, vol. 15, no. 19, pp. 4264–4272, Sep. 1976.
 K. Takeda et al. J. Colloid Interface Sci., vol. 154, no. 2, pp. 385–392, Dec. 1992.

![](_page_0_Picture_41.jpeg)

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